

# British Association of Sexual Health and HIV National guideline for the management of infection with *Mycoplasma genitalium*, 2025

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## Abstract

This guideline provides details on the pathology and clinical features of *Mycoplasma genitalium* infection and makes recommendations for diagnostic tests, treatment regimens and the health promotion principles needed for the effective management of infection, in people aged 16 years or older attending sexual health services. The guideline is primarily aimed at level 3 sexual health services in the UK, although it could also serve as a reference guide for sexually transmitted infections services at other levels. It is updated from the previous guideline published in 2018.

## Keywords

Non-gonococcal urethritis, urethritis, *Mycoplasma genitalium*, antimicrobial resistance

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## What is new in the 2025 guideline?

- An updated description of the complications associated with *M. genitalium* infection is included.
- Indications for *M. genitalium* testing are now based on specific syndromes (instead of individual symptoms) and/or risk factors. There is reiteration that asymptomatic screening should not be undertaken.
- The evidence base for recommended treatments is expanded in light of case reports, clinical series and systematic reviews published since the last version of this guideline.
- Clinicians are advised to counsel patients on the risks of damage to tendons, muscles, joints and nerves, as well as significant psychiatric side effects including suicidal intent when moxifloxacin is used.
- A test of cure (TOC) is no longer recommended for all patients with confirmed *M. genitalium* infection. Re-testing can be undertaken in individuals who remain symptomatic after treatment and when there is suspicion of persistent infection.

## Introduction and methodology

### Objectives

The guideline is primarily aimed at health care professionals seeing people aged 16 years or older in departments offering

specialist level 3 care in sexually transmitted infection (STI) management within the United Kingdom. However, the principles of the recommendations can be observed across other levels of STI care providers. Non-specialist services may need to develop local referral pathways.

This guideline provides details on the pathology and clinical features of *Mycoplasma genitalium* infection, and offers recommendations on the diagnostic tests, treatment regimens and

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health promotion principles needed for the effective management of infection. Whilst the guideline is intended to summarise recommendations for best practice according to current available evidence, it is acknowledged that not all clinics will have ready access to *M. genitalium* detection and resistance testing because of logistical and cost issues.

### Search strategy and methods

This guideline was produced according to specifications set out in the British Association for Sexual Health and HIV (BASHH) Clinical Effectiveness Group (CEG) 'framework for guideline development and assessment' (2015, updated 2020) accessed at [https://www.bashh.org/\\_userfiles/pages/files/resources/2020\\_guidelines\\_framework.pdf](https://www.bashh.org/_userfiles/pages/files/resources/2020_guidelines_framework.pdf).

This guideline has been updated by reviewing the previous BASHH national guideline for the management of infection with *M. genitalium* (2018)<sup>1</sup> and conducting a comprehensive literature search of publications from January 2017 to April 2024. Conference abstracts were updated when the published manuscript became available. The writing group formulated eleven 'PICO' questions (see Appendix 1) addressing the patient problem or population (P), intervention (I) (or aetiology/diagnosis/frequency/prognosis), comparison (C) and outcome(s) (O), which formed the basis for the literature search and article identification. The search was conducted using Medline, Embase, Cochrane library and National Health Service (NHS) Evidence databases and the broad search term '*Mycoplasma genitalium*'. All entries in the English language were reviewed, and if relevant the full text obtained. Age, country, and study design limits were included in the PICO criteria. 'Grey literature' included conference abstracts from International Union against Sexually Transmitted Infection (IUSTI), BASHH, British HIV Association (BHIVA), Interscience Conference of Antimicrobial Agents and Chemotherapy (ICAAC), Australasian Society for HIV, Viral Hepatitis and Sexual Health Medicine (ASHM) and European Society of Clinical Microbiology and Infectious Diseases (ECCMID) scientific meetings in the last 3 years.

The Grading of Recommendations, Assessment, Development, and Evaluations (GRADE) system was used to assess the evidence and make recommendations (see Appendix 2) and consensus was reached on all recommendations.

### Equality impact assessment

An assessment of the guideline and its recommendations was undertaken to ensure the principles of equality and diversity were adhered to and is available in Appendix 3.

BASHH has adopted an anatomical approach without assuming gender in the majority of guidelines and uses gender terminology in line with BASHH 'Sexual health standards for trans, including non-binary, people'.

The terminology of 'men' and 'women' has been retained in a few instances relating to other guidelines, e.g., human papillomavirus vaccination and female genital mutilation.

### Stakeholder involvement, piloting and feedback

The first draft was produced by the writing group and then circulated to the BASHH Clinical Effectiveness Group (CEG) for review using the the Appraisal of Guidelines, Research and Evaluation (AGREE) appraisal tool Appendix 4. The second draft of the guideline was posted on the BASHH website for wider consultation (2 months) and any comments received during the consultation period were reviewed by the authors and acted on appropriately. The document was also reviewed by a patient representative, target users and the public panel of BASHH, and their feedback was considered by the authors and used to inform the guideline. The final draft was presented to the CEG for review and piloting in sexual health clinics.

Once the guideline is published, the CEG will keep it under review should critical new evidence become available that affects the current recommendations. The guideline will be formally reviewed and updated, if necessary, every 5 years.

## Epidemiology

### Prevalence in the general population

In England, prevalence estimates of *M. genitalium* infection range from 1% to 2% in the general population, being slightly higher in women than men.<sup>2</sup> Amongst STI clinic attendees, prevalence ranges are higher, from 4 to 38%.<sup>3-6</sup>

### Risk factors for infection

Similar to *C. trachomatis*, risk factors for *M. genitalium* infection include younger age, non-white ethnicity, smoking, and increasing number of sexual partners.<sup>2</sup> However, the prevalence of *M. genitalium* infection appears to peak later than that for *C. trachomatis*, particularly in men, and to remain higher in older age groups. The use of pre-exposure prophylaxis (PrEP) has been associated with an increased risk of *M. genitalium* infection. In a recent meta-analysis, the prevalence of *M. genitalium* in individuals using PrEP was higher (16.7%) compared to non-users.<sup>7</sup>

### Sexual transmission

Transmission is primarily by genital-genital contact, but transmission by penile-anal contact has also been established following detection of organism in the anorectal compartment.<sup>8-11</sup> As carriage in the oro-pharynx is uncommon, the relative contribution of oral sex is likely to be very small.<sup>12-14</sup> The risk of transmission per coital act has

yet to be determined but is likely to be less than that for *Chlamydia trachomatis*.<sup>14</sup>

### Coinfection with other sexually transmitted infections

As *M. genitalium* is transmitted through sexual contact, co-infections with other bacterial STIs such as *C. trachomatis* and *Neisseria gonorrhoeae* are common.<sup>15,16</sup> An association between *M. genitalium* and HIV transmission and acquisition is biologically plausible and supported by some studies in Africa.<sup>6,17,18</sup>

## Clinical features

### Asymptomatic infection

The evidence suggests that the majority of people infected with *M. genitalium* in the genital tract do not develop disease.<sup>14,19,20</sup> There are limited data to suggest that infection can be cleared spontaneously within a few months although persistence for up to 2 years is also reported.<sup>20,21</sup> Current treatments are imperfect and associated with the development of antimicrobial resistance (AMR).<sup>22,23</sup> There is no evidence that screening asymptomatic individuals will be of benefit, and indeed is likely to do harm at a population level.<sup>24</sup>

### Symptomatic infection

#### Signs and symptoms

**Penile urethral infection.** *M. genitalium* infection is unequivocally and strongly associated with non-gonococcal urethritis (NGU). Typically, the prevalence of *M. genitalium* in men with NGU is 15%–25% and in men with non-chlamydial nongonococcal urethritis (NCNGU) is 10%–35%.<sup>25</sup> *M. genitalium* is also associated with persistent and recurrent urethritis, where up to 40% of affected men may have *M. genitalium* detected.<sup>26</sup>

The clinical presentation of *M. genitalium* urethritis is similar to other causes and thus clinical features of acute symptomatic NGU cannot be used to determine the infective aetiology.<sup>14,27–31</sup> The commonest symptoms are discharge and urethral irritation. Although the proportion of infected men that develop symptoms is unknown, this is likely to be <10%.<sup>14</sup>

**Endocervical infection.** Several studies support an association of *M. genitalium* infection in cisgender women with post-coital bleeding and cervicitis.<sup>32–34</sup> Although other genital symptoms including vaginal discharge, odour, itch, dyspareunia and/or bleeding may be apparent during infection, studies have found no association with *M. genitalium* and their presence alone is not indicative of a testing need.<sup>34</sup>

**Rectal infection.** *M. genitalium* is commonly found in rectal samples from gay, bisexual and other men who have sex with men (GBMSM).<sup>35,36</sup> While some studies

show no association with symptoms,<sup>9</sup> a recent systematic review did demonstrate an overall association of *M. genitalium* infection with proctitis.<sup>37</sup> There appears to be a correlation between organism load and symptoms suggesting causation.<sup>8,9,38</sup> To avoid overtreatment, and given that proctitis caused by *C. trachomatis* or *N. gonorrhoeae* mono-infection is more likely than proctitis caused by *M. genitalium*, routinely testing for *M. genitalium* in proctitis presentations is not recommended, except in specific circumstances (see Indications for Testing).<sup>39,40</sup>

**Pharyngeal infection.** A low prevalence of *M. genitalium* in pharynx is consistently reported, suggesting that this is not an important reservoir in the transmission of infection.<sup>36,37</sup>

**Ocular infection.** Ocular infection with *M. genitalium* can lead to conjunctivitis.<sup>41</sup> There are no reports of neonatal conjunctivitis when born to mothers with *M. genitalium* infection.

### Complications

**Pelvic inflammatory disease.** *M. genitalium* is linked aetiologically to pelvic inflammatory disease (PID) and accounts for 9–13% of cases of PID.<sup>42–44</sup> A meta-analysis in 2015 demonstrated significant associations between the presence of *M. genitalium* and cervicitis (pooled OR 1.66) and PID (pooled OR 2.14).<sup>33</sup>

The organism has been shown to ascend from the lower to upper female genital tract, has been detected frequently from endometrial biopsies in individuals with PID independent of *C. trachomatis* and *N. gonorrhoeae*<sup>45,46</sup> and can cause epithelial ciliary damage in human fallopian tube culture. However, an association with tubal factor infertility has not yet been demonstrated and conducting studies to determine this will be difficult.<sup>25,47,48</sup>

It is estimated that 4.9% of individuals with *M. genitalium* in the female genital tract will progress to PID (compared to 14.4% of those with *C. trachomatis* infection).<sup>44</sup>

**Epididymo-orchitis.** A small number of studies suggest an association of *M. genitalium* infection with epididymo-orchitis.<sup>49,50</sup> Due to insufficient data, routinely testing for *M. genitalium* in epididymo-orchitis presentations is not recommended, except in specific circumstances (see Indications for Testing).

**Prostatitis.** Although *M. genitalium* has been detected in people with prostatitis and chronic pelvic pain syndrome,<sup>51,52</sup> current data are lacking to support a clear association with prostatitis.

**Sero-negative reactive arthritis.** Case reports and case series indicate a possible association between *M. genitalium* and sexually acquired reactive arthritis.<sup>14,25,53,54</sup>

## Pregnancy

A small number of studies have examined outcomes of *M. genitalium* in pregnancy. There is a possible increased risk of preterm birth (OR 1.91) but no association with spontaneous abortion.<sup>55</sup> There are insufficient data for premature rupture of membranes, low birth weight and perinatal death.

## Indications for testing

### Based on syndromes

- It is recommended to test for *M. genitalium* infection in people with NGU. (Grade 1B)
- It is recommended to test for *M. genitalium* infection in people with signs and symptoms suggestive of PID. (Grade 1B)

Although *M. genitalium* may present with a range of symptoms (see Clinical features), it is often non-pathological and not the causative organism. Routine testing in these situations would result in overdiagnosis and overtreatment. Therefore, it is only recommended to test for *M. genitalium* in the following syndromes if symptoms persist after other infections have been ruled out and the index of suspicion is high:

- People with signs or symptoms of mucopurulent cervicitis, particularly post-coital bleeding (Grade 2B);
- People with epididymo-orchitis (Grade 2D);
- People with sexually-acquired proctitis (Grade 2D).

An alternative strategy is to test for *M. genitalium* based on persistent symptoms and where *C. trachomatis* and *N. gonorrhoeae* screening is negative. However, this is not a preferable approach because it results in delayed identification of infection and treatment.

### Based on risk factors

It is recommended to test for *M. genitalium* infection in current sexual partners of *M. genitalium*-infected individuals (see Tracing and Treatment of Contacts) (Grade 1D).

### Asymptomatic individuals

There are currently insufficient data to recommend routine screening for *M. genitalium* infection in asymptomatic individuals. Newer multiple pathogen assays which include the detection of *M. genitalium* are being used by some providers in STI screening of asymptomatic individuals. This is not recommended.<sup>56</sup> Asymptomatic individuals with confirmed *C. trachomatis* and/or *N. gonorrhoeae* infection should not routinely be tested for *M. genitalium*.

## Diagnosis

### Diagnostic tests

*M. genitalium* has fastidious nutritional requirements and is extremely slow growing therefore culture is not appropriate for diagnosis. Nucleic acid amplification tests (NAATs) that detect *M. genitalium*-specific DNA or RNA in clinical specimens are the only useful diagnostic method.

It is recommended to test all *M. genitalium*-positive specimens for macrolide resistance associated mutations (MRAMs) using the commercial assays available in the UK. Commercial assays available for the detection of fluoroquinolone resistance associated mutations are not widespread but are becoming available. The UK Health Security Agency (UKHSA) STI Reference Laboratory offers a fluoroquinolone susceptibility genotyping assay for cases of treatment failure.

### Specimen collection

**Penile urethra.** Most studies have found that first void urine and urethral swabs have good sensitivity (>90%) for the detection of *M. genitalium*.<sup>57–62</sup> There are conflicting data for self-taken meatal swabs, with sensitivity estimates of 79–88%.<sup>62,63</sup>

**Female genital tract.** Vulvovaginal swabs are the most sensitive specimen (sensitivity >98%). Self-taken specimens have comparable diagnostic accuracy to clinician-taken specimens.<sup>62,64</sup> The sensitivity of endocervical swabs and urine is lower (74–81% and 58–86.4%, respectively).<sup>57,62,65–67</sup>

**Non-genital sampling.** Rectal swabs are suitable for sampling and can be self-taken.

Pharyngeal *M. genitalium* is not thought to be clinically relevant and sampling is not recommended.

**Considerations for people following gender reassignment surgery.** There are a paucity of data concerning *M. genitalium* infection in individuals following gender reassignment surgery. It is therefore difficult to recommend an optimal specimen type, but this should be guided by sexual history and symptoms.

### Recommendations

- It is recommended to use first void urine as the specimen of choice in cisgender men (Grade 1C).
- It is recommended to use vaginal swabs (clinician- or self-taken) as the specimen of choice in cisgender women (Grade 1C).
- It is recommended, where possible, to test all *M. genitalium*-positive specimens for the presence of MRAMs (Grade 1B).

## Window period

There are no data on the incubation period for *M. genitalium*, or on the likely window period before a laboratory test becomes reliably positive. However, it is likely that sensitive tests will detect early infection.

## Management

### General advice

Patients should be given a detailed explanation of their condition, reinforced with clear and accurate written information. A patient information leaflet for *M. genitalium* can be found on the resource page of the BASHH website.<sup>68</sup> This will be updated when new guidance is published or new information becomes available.

Patients should be advised to abstain from sexual intercourse until 14 days after the start of treatment, and until symptoms have resolved. Where azithromycin has been used this is especially important because of its long half-life and is likely to reduce the risk of selecting/inducing macrolide resistance if the patient is re-exposed to *M. genitalium*.

## Treatment

### Recommended regimens

#### Treatment of uncomplicated infection (urethritis, cervicitis)

**Azithromycin and macrolide resistance.** Eradication rates of *M. genitalium* following treatment with macrolides have declined globally in recent years, and rates of genotypic resistance range from 30% to 100% regionally.<sup>69</sup> A recent systematic review and meta-analysis of 166 studies, including 22,974 samples across 41 countries demonstrated a global point prevalence of genotypic macrolide resistance of 33%.<sup>70</sup> Data from UK surveillance studies estimate macrolide resistance in the UK to be around 62.2% with the highest proportions of resistance seen in GBMSM. Use of azithromycin 2 g over 3 days in the absence of MRAMS is associated with a high rate of treatment success (94%).<sup>71</sup> Although there is no direct evidence to suggest that this regimen is superior in efficacy to longer courses of azithromycin, the risk of developing macrolide resistance is likely to be lower whilst offering a shorter and well tolerated course.

Access to prompt MRAM testing varies across the UK centres but is a useful tool to guide appropriate antibiotic therapy and improve treatment outcomes where available in a timely manner.

Even where an organism is known to be initially macrolide-sensitive, an azithromycin regimen should not be repeated following treatment failure because it is likely that resistance has developed on treatment.

**Doxycycline.** Although doxycycline as monotherapy has poor efficacy and eradication rates are low at about 30%–40%, studies suggest AMR-guided therapy, with doxycycline initially followed by azithromycin or moxifloxacin, based on the presence or absence of MRAMs, may increase cure rates after appropriate first-line therapy to over 92%.<sup>72,73</sup> This is biologically plausible, as doxycycline reduces the organism load and hence the risk of pre-existing genotypic determinants of resistance being present.

**Moxifloxacin and quinolone resistance.** Moxifloxacin still has good efficacy in Europe<sup>29,74</sup> although resistance is increasing in the Asia-Pacific region where its use is greater.<sup>75</sup> A recent systematic review and meta-analysis of 139 studies has demonstrated that global fluoroquinolone resistance rose from 10.5% (7.0–14.5%) in 2015–2017 to 17.0% (11.2–23.5%) in 2018–2020 and from 8% to 12% in non-Nordic European countries specifically.<sup>70</sup>

Genotypic determinants of quinolone resistance are complex and incompletely understood. Mutations more commonly arise in the *parC* gene than in the *gyrA* gene. Mutations in *parC* gene may confer reduced susceptibility to moxifloxacin, and the occurrence of mutations in both genes is a strong predictor of treatment failure.<sup>76</sup> Recent enhanced surveillance data of *M. genitalium* samples in the UK detected fluoroquinolone resistance in 12.1% specimens with *parC* sequencing, and MRAM/*parC* dual class mutations in 10.1%.<sup>71</sup>

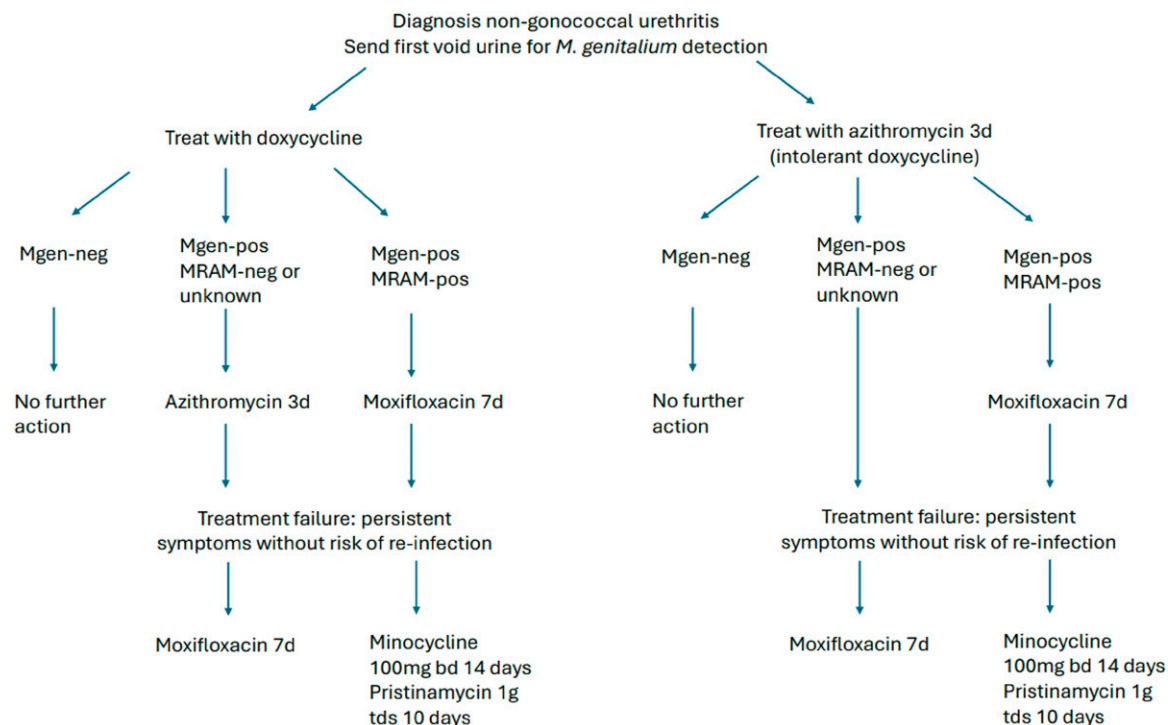
Using moxifloxacin first-line in all cases of *M. genitalium* infections is not recommended because future therapeutic options are limited. Furthermore, the 2024 update on the use of fluoroquinolones published by the Medicines and Healthcare Products Regulation Agency (MHRA) has restricted the systematic use of this class of antibiotics to only situations where no alternative antibiotics are suitable.<sup>77</sup> As treatment options for *M. genitalium* are restricted in the presence of MRAMs, it remains appropriate to use moxifloxacin as an alternative first-line therapy, though patients require counselling on the risks of damage to tendons, muscles, joints and nerves, as well as significant psychiatric side effects including suicidal intent. The MHRA has produced patient information leaflets to provide to patients commencing quinolone therapy which are accessible via the MHRA website.

A meta-analysis reported no significant difference between moxifloxacin regimens of 7- and 10-days' duration,<sup>78</sup> and considering the MHRA guidance, a 7-day course is recommended in uncomplicated infections.

See Figure 1 a suggested treatment pathway for men presenting with NGU who subsequently test positive for *M. genitalium*.

#### Recommended regimens (uncomplicated infections):

- Doxycycline 100 mg twice daily (bd) for 7 days followed by azithromycin 1 g orally as a single dose then 500 mg



**Figure 1.** Suggested treatment pathway for men presenting with non-gonococcal urethritis who subsequently test positive for *Mycoplasma genitalium*. Azithromycin 3d should be started within 2 weeks of finishing doxycycline. Azithromycin 3d: azithromycin 1 g, then 500 mg od for 2 days; bd: twice daily; d: days; Doxycycline: doxycycline 100 mg bd for 7 days; Mgen: *Mycoplasma genitalium*; Moxifloxacin 7d: moxifloxacin 400 mg bd for 7 days; MRAM: macrolide resistance associated mutation; neg: negative; pos: positive; tds: three times a day.

orally od for 2 days\* where the organism is known to be macrolide-sensitive or where resistance status is unknown (Grade 1C).

- Doxycycline 100 mg bd for 7 days followed by moxifloxacin 400 mg orally od for 7 days if organism is known to be macrolide-resistant or where treatment with azithromycin has failed\*\* (Grade 1C).

\* Given that many individuals will have had doxycycline as first-line therapy for uncomplicated infectious syndromes, a repeat course is unnecessary once the *M. genitalium* positive result is known. Azithromycin and/or moxifloxacin should be given immediately after doxycycline, and ideally within 2 weeks of completing doxycycline. If this is not possible, the course of doxycycline should be repeated prior to giving azithromycin or moxifloxacin.

\*\* Treatment failure is defined as persistent symptoms following treatment, or a positive test in the presence of symptoms taken 5 weeks post-treatment (if indicated – see Section Follow Up).

In situations where clinical cure is achieved prior to confirmation of a positive *M. genitalium* result, it may be pragmatic to take a watch and wait approach rather than offering further antimicrobial therapy.

**Alternative regimens.** In cases of failure or intolerance of recommended regimens, there are some observational data to support alternative regimens.

In one study, treatment with minocycline 100 mg od for 14 days was generally well-tolerated and cured approximately 67% of people who experienced treatment failure with a previous regimen.<sup>79</sup> The minimum inhibitory concentrations (MICs) of minocycline for reference strains of *M. genitalium* have been shown to be lower than those of doxycycline, thus greater effectiveness might be inferred (but, as with doxycycline, MICs of tetracyclines may not predict treatment outcomes).<sup>80</sup>

Pristinamycin is a streptogramin antibiotic. It is available on a named patient basis in England for several indications, including treatment of *M. genitalium* infection failing recommended regimens.

An observational analysis of 114 people with macrolide-resistant *M. genitalium* infection showed 75% cure when treated with pristinamycin.<sup>81</sup> For infections with higher pre-treatment bacterial load, treatment was more likely to fail. Hence to improve chances of organism clearance, pre-treatment with doxycycline is advised. Patients should be warned of gastrointestinal side effects.

A recent observational study investigated the tolerability and efficacy of combination therapy of minocycline 100 mg

twice daily and metronidazole 400 mg twice daily for 14 days (with or without pre-treatment with doxycycline) in individuals with macrolide-resistant *M. genitalium* in whom fluoroquinolones had failed or were not advised.<sup>82</sup> In patients receiving pre-treatment with doxycycline, the microbiological cure rate was 90.3% ( $n = 28/31$ ) compared to 76.7% ( $n = 56/73$ ) in those who had not. Central nervous system and gastrointestinal side effects were commonly reported, however.

In cases of treatment failure with, or intolerance of first- and second-line regimens (including doxycycline, azithromycin, moxifloxacin), it is recommended to use:

- Minocycline 100 mg orally bd for 14 days (Grade 1C).
- Doxycycline 100 mg bd for 7 days followed by pristinamycin 1000 mg orally three times daily for 10 days (Grade 1C).

**Treatment of complicated urogenital infection (PID, epididymo-orchitis).** There are few studies examining the efficacy of extended azithromycin and moxifloxacin regimens in the treatment of PID and epididymo-orchitis caused by *M. genitalium*. Given the need for prompt and effective treatment in complex STI syndromes, individuals with confirmed *M. genitalium* infection should be given moxifloxacin as a 14-day regimen.<sup>78</sup> Moxifloxacin is the currently the most efficacious agent against *M. genitalium* and an extended regimen has the highest chance of eradication. Individuals should be switched to moxifloxacin (if another regimen has been started) as soon as possible after *M. genitalium* is confirmed; doxycycline pre-treatment is not necessary in this situation. However, if treatment for PID has been completed and symptoms have resolved by the time of the *M. genitalium* result then it is pragmatic to review symptoms rather than re-treat all with moxifloxacin.

#### **Recommended regimens (complicated infection)**

- Moxifloxacin 400 mg orally od for 14 days (Grade 1C).

#### **Alternative regimens**

- Azithromycin 1 g orally stat followed by 500 mg orally od for 4 days (Grade 2A).

### **Pregnancy and breast-/chest-feeding**

**Pregnancy.** Where possible, it is advisable to delay treatment until after pregnancy.<sup>83</sup> Clinicians should have an informed discussion with the pregnant person about the risks associated with treatment during pregnancy and the low risk of adverse pregnancy outcomes associated with *M. genitalium* infection. Condom use during the remainder of pregnancy is advised to reduce the risk of onward transmission of infection to sexual partners.

Treatment options in pregnancy are constrained by limited available data. There is some evidence that the use of azithromycin during pregnancy is unlikely to increase the risk of birth defects or adverse pregnancy outcomes.<sup>84–86</sup> It is recommended a longer course of azithromycin to treat PID in pregnant people. This should be 1 g stat followed by 500 mg od for 4 days (Grade 1A).

Although the BNF recommendation is to avoid tetracyclines at all stages of pregnancy, BASHH supports the use of doxycycline in pregnant people only when the full course of treatment can be completed prior to 15 weeks gestation.<sup>87</sup> (Grade 2D)

The use of moxifloxacin in pregnancy is contraindicated.<sup>88</sup> Minocycline use during pregnancy is contraindicated.<sup>89</sup>

Pristinamycin is considered safe in pregnancy, given its widespread use in Europe, but the writing group acknowledges that there are a lack of data supporting this.<sup>81</sup>

**Breast-/Chest-feeding.** The level of antibiotic detected in breast milk of those taking azithromycin is very low and the systemic exposure in infants does not exceed that observed when azithromycin is administered for treatment.<sup>90</sup> Therefore, the risk for the infant is considered to be low. Infants should be monitored for possible side effects on the gastrointestinal flora including diarrhoea and candidiasis. A large cohort study found an association between the maternal use of macrolides during the first 13 days after delivery and an increased risk of pyloric stenosis in breastfed infants.<sup>91</sup> Where possible, exposure of neonates to macrolides should be minimised and precautionary monitoring undertaken with use. Doxycycline is excreted into breast milk in very small amounts and long term use is contraindicated due to the risk of tooth discolouration and effects on bone growth.<sup>92</sup> However, shorter courses (<21 days) are unlikely to cause harm because drug absorption by the infant is inhibited by calcium in breastmilk, and use of 7 days doxycycline as part of dual treatment for *M. genitalium* is acceptable.<sup>93</sup> Moxifloxacin is contraindicated during breast/chest-feeding.<sup>88</sup> Use of pristinamycin during breast/chest feeding has been associated with side effects in the baby. Due to a lack of data overall, its use is not recommended.<sup>94</sup>

### **People living with HIV taking antiretroviral therapy**

Treatment of *M. genitalium* in people living with HIV is the same as that for people living without HIV.

### **Adverse effects**

Azithromycin, doxycycline, minocycline, moxifloxacin and pristinamycin can all cause gastrointestinal problems including nausea, but symptoms are most frequently reported with doxycycline and azithromycin doses over 1 g.<sup>95</sup>

Caution should be taken when prescribing azithromycin or moxifloxacin to patients already on medications which may prolong the QT interval.

Minocycline has been associated with early onset toxicity reactions, hypersensitivity, autoimmune disorders, skin pigmentation and hepatitis. The risk of adverse effects increases with the duration of use and cumulative dose.

The MHRA has strengthened the recommendation on restricting the use of fluoroquinolone antibiotics following a review of their disabling and potentially long-lasting side effects.<sup>77</sup> Healthcare professionals should advise patients to stop treatment with a fluoroquinolone antibiotic at the first sign of side effects including tendonitis or tendon rupture, muscle pain/weakness, joint pain/swelling, peripheral neuropathy and central nervous system/psychiatric effects. Hepatotoxicity has been reported but is very rare (<1/10,000).

## Follow-up

Persistence of *M. genitalium* has been demonstrated in the absence of symptoms in men treated for NGU.<sup>96,97</sup> This occurs in about 10%–20% of men treated with doxycycline but is not associated with the development of AMR.<sup>14,97</sup> Persistence of *M. genitalium* following treatment with azithromycin and moxifloxacin is strongly associated with AMR.<sup>96,97</sup> Choice in efficacious therapeutic agents subsequent to this is limited and achievement of microbiological cure is not always possible. Thus, a more pragmatic aim for successful treatment should be the resolution of symptoms.

If symptoms persist following treatment with a regimen deemed efficacious, the risk of re-infection should be excluded and compliance with medication should be verified. Re-testing can be considered to determine whether symptoms are specific to *M. genitalium* and thus whether further treatment is warranted. The optimal time to perform this test after treatment has not been determined, but data suggest that very early testing after treatment when DNA load is low can give false negative results.<sup>98</sup>

It is advisable that:

- A test of cure (TOC) is not required where there is resolution of symptoms; clinical rather than microbiological cure should be pursued in all people (Grade 1D);
- Re-testing can be performed in individuals who remain symptomatic after treatment when there is suspicion of persistent infection (Grade 1D)

## Tracing and treatment of contacts

Patients should be informed of the importance of partner notification and supported to do this by appropriately trained professionals.

Among heterosexual contacts, women were twice as likely as men to be infected with *M. genitalium*, after

adjusting for condom use and nature of relationship.<sup>99</sup> Among those patients reporting sexual contact with an *M. genitalium*-infected individuals, 48.2% of women, 31.0% of men who have sex with women, and 41.7% of GBMSM were infected.<sup>99,100</sup>

Whilst acknowledging this, the primary purpose of partner notification is to reduce the risk of re-infection to the symptomatic index patient.

Only current partner(s) (including non-regular partners where there is likely to be further sexual contact) should be tested, and treated only if positive. Sampling the rectum in contacts of index patients may be indicated by sexual history; we do not recommend sampling the pharynx. Partners should be given the same antibiotic as the index patient unless there is available resistance information to suggest otherwise (Grade 1D).

## Auditable outcome measures

- All cases of *M. genitalium* infection should prompt a Sexual Health and HIV Activity Property Type (SHHAPT) code 'C16' submission to Genitourinary Medicine Clinic Activity Dataset (GUMCAD, performance standard 97%).
- Cases of microbiologically confirmed treatment failure with persistent symptoms following moxifloxacin therapy should be reported to UKHSA at: <https://www.gov.uk/government/organisations/uk-health-security-agency> (performance standard 97%).
- Individuals presenting with *M. genitalium* infection should be provided with written information about their diagnosis and management (performance standard 97%); e.g., patient information leaflets on *M. genitalium* from BASHH.<sup>68</sup>
- Partner notification should be performed according to this guideline and documented according to the BASHH statement on partner notification for STIs (performance standard 97%).<sup>101</sup>

## Recommendations for further research

Longitudinal data are needed to determine natural history and time to clearance of organism, the longer-term effects of *M. genitalium* on the genital tract and clearer associations with reproductive health consequences.

Randomised controlled trials in clinical settings examining the efficacy of new antibiotic regimens are urgently needed to increase the therapeutic options available to manage *M. genitalium*. Whilst RCTs for newer drugs have shown some promise with respect to *M. genitalium*, approval for these newer agents is not yet available.

## Qualifying statement

The recommendations in this guideline may not be appropriate for use in all clinical situations. Decisions to

follow these recommendations must be based on professional clinical judgement, consideration of individual patient circumstances and available resources.

All possible care has been undertaken to ensure specification of the correct dosage of medication and route of administration. However, it remains the responsibility of the prescribing clinician to ensure the accuracy and appropriateness of the medication they prescribe.

## Review arrangements

An author group will be invited by the BASHH CEG to review and revise the guideline in 2030 using the BASHH framework for guideline development. However, addenda may be issued sooner than 2030, particularly if relevant new data are available relating to testing or treatment options.

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## Membership of the clinical effectiveness group

Three members of the writing group are current members of the BASHH CEG. Current membership of the BASHH CEG is available at [https://www.bashh.org/professionals/bashh\\_groups/39/clinical\\_effectiveness\\_group/public](https://www.bashh.org/professionals/bashh_groups/39/clinical_effectiveness_group/public)

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## Supplemental Material

Supplemental material for this article is available online.

## References

1. Soni S, Horner P, Rayment M, et al. British Association for Sexual Health and HIV national guideline for the management of infection with *Mycoplasma genitalium* (2018). *Int J STD AIDS* 2019; 30(10): 938–950.
2. Sonnenberg P, Ison CA, Clifton S, et al. Epidemiology of *Mycoplasma genitalium* in British men and women aged 16–44 years: evidence from the third national survey of sexual attitudes and lifestyles (Natsal-3). *Int J Epidemiol* 2015; 44(6): 1982–1994.
3. Getman D, Jiang A, O'Donnell M, et al. *Mycoplasma genitalium* prevalence, coinfection, and macrolide antibiotic resistance frequency in a multicenter clinical study cohort in the United States. *J Clin Microbiol* 2016; 54(9): 2278–2283.
4. Hetem DJ, Kuizenga Wessel S, Bruisten SM, et al. High prevalence and resistance rates of *Mycoplasma genitalium* among patients visiting two sexually transmitted infection clinics in The Netherlands. *Int J STD AIDS* 2021; 32(9): 837–844.
5. le Roux MC and Hoosen AA. Quantitative real-time polymerase chain reaction for the diagnosis of *Mycoplasma genitalium* infection in South African men with and without symptoms of urethritis. *Sex Transm Dis* 2017; 44(1): 17–20.
6. Napierala M, Munson E, Wenten D, et al. Detection of *Mycoplasma genitalium* from male primary urine specimens: an epidemiologic dichotomy with *Trichomonas vaginalis*. *Diagn Microbiol Infect Dis* 2015; 82(3): 194–198.
7. Sokoll PR, Migliavaca CB, Siebert U, et al. Prevalence of *Mycoplasma genitalium* infection among HIV PrEP users: a systematic review and meta-analysis. *Sex Transm Infect* 2023; 99(5): 351–359.
8. Bissessor M, Tabrizi SN, Bradshaw CS, et al. The contribution of *Mycoplasma genitalium* to the aetiology of sexually acquired infectious proctitis in men who have sex with men. *Clin Microbiol Infect* 2016; 22(3): 260–265.
9. Soni S, Alexander S, Verlander N, et al. The prevalence of urethral and rectal *Mycoplasma genitalium* and its associations in men who have sex with men attending a genitourinary medicine clinic. *Sex Transm Infect* 2010; 86(1): 21–24.
10. Edlund M, Blaxhult A and Bratt G. The spread of *Mycoplasma genitalium* among men who have sex with men. *Int J STD AIDS* 2012; 23(6): 455–456.
11. Bradley I, Varma R, Knight V, et al. Prevalence of rectal *Mycoplasma genitalium* and macrolide resistance in men who have sex with men attending Sydney Sexual Health Centre. *Sex Health* 2020; 17(2): 114–120.
12. Bradshaw CS, Fairley CK, Lister NA, et al. *Mycoplasma genitalium* in men who have sex with men at male-only saunas. *Sex Transm Infect* 2009; 85(6): 432–435.
13. Deguchi T, Yasuda M, Yokoi S, et al. Failure to detect *Mycoplasma genitalium* in the pharynges of female sex workers in Japan. *J Infect Chemother* 2009; 15(6): 410–413.
14. Horner PJ and Martin DH. *Mycoplasma genitalium* infection in men. *J Infect Dis* 2017; 216(suppl 2): S396–S405.
15. Harrison SA, Olson KM, Ratliff AE, et al. *Mycoplasma genitalium* coinfection in women with Chlamydia trachomatis infection. *Sex Transm Dis* 2019; 46(10): e101–e104.
16. Richardson D, Lewis DA, Jeoffreys NJ, et al. *Mycoplasma genitalium* coinfection in men with symptomatic gonococcal urethritis. *Sex Transm Infect* 2021; 97(5): 363–367.

17. Vandepitte J, Weiss HA, Kyakuwa N, et al. Natural history of *Mycoplasma genitalium* infection in a cohort of female sex workers in Kampala, Uganda. *Sex Transm Dis* 2013; 40(5): 422–427.
18. Mavedzenge SN, Van Der Pol B, Weiss HA, et al. The association between *Mycoplasma genitalium* and HIV-1 acquisition in African women. *AIDS* 2012; 26(5): 617–624.
19. Golden MR, Workowski KA and Bolan G. Developing a public health response to *Mycoplasma genitalium*. *J Infect Dis* 2017; 216(suppl\_2): S420–S426.
20. Oakeshott P, Aghaizu A, Hay P, et al. Is *Mycoplasma genitalium* in women the “New Chlamydia?” A community-based prospective cohort study. *Clin Infect Dis* 2010; 51(10): 1160–1166.
21. Balkus JE, Manhart LE, Jensen JS, et al. *Mycoplasma genitalium* infection in Kenyan and US women. *Sex Transm Dis* 2018; 45(8): 514–521.
22. Bradshaw CS, Jensen JS and Waites KB. New horizons in *Mycoplasma genitalium* treatment. *J Infect Dis* 2017; 216(suppl\_2): S412–S419.
23. Horner P, Ingle SM, Garrett F, et al. Which azithromycin regimen should be used for treating *Mycoplasma genitalium*? A meta-analysis. *Sex Transm Infect* 2018; 94(1): 14–20.
24. Bradshaw CS, Horner PJ, Jensen JS, et al. Syndromic management of STIs and the threat of untreatable *Mycoplasma genitalium*. *Lancet Infect Dis* 2018; 18(3): 251–252.
25. Taylor-Robinson D and Jensen JS. *Mycoplasma genitalium*: from Chrysalis to multicolored butterfly. *Clin Microbiol Rev* 2011; 24(3): 498–514.
26. Wikstrom A and Jensen JS. *Mycoplasma genitalium*: a common cause of persistent urethritis among men treated with doxycycline. *Sex Transm Infect* 2006; 82(4): 276–279.
27. Falk L, Fredlund H and Jensen JS. Symptomatic urethritis is more prevalent in men infected with *Mycoplasma genitalium* than with *Chlamydia trachomatis*. *Sex Transm Infect* 2004; 80(4): 289–293.
28. Jensen JS, Orsum R, Dohn B, et al. *Mycoplasma genitalium*: a cause of male urethritis? *Genitourin Med* 1993; 69(4): 265–269.
29. Anagrus C, Loré B and Jensen JS. Treatment of *Mycoplasma genitalium*. Observations from a Swedish STD clinic. *PLoS One* 2013; 8(4): e61481.
30. Wetmore CM, Manhart LE, Lowens MS, et al. Demographic, behavioral, and clinical characteristics of men with non-gonococcal urethritis differ by etiology: a case-comparison study. *Sex Transm Dis* 2011; 38(3): 180–186.
31. Ito S, Hanaoka N, Shimuta K, et al. Male non-gonococcal urethritis: from microbiological etiologies to demographic and clinical features. *Int J Urol* 2016; 23(4): 325–331.
32. Manhart LE, Critchlow CW, Holmes KK, et al. Mucopurulent cervicitis and *Mycoplasma genitalium*. *J Infect Dis* 2003; 187(4): 650–657.
33. Lis R, Rowhani-Rahbar A and Manhart LE. *Mycoplasma genitalium* infection and female reproductive tract disease: a meta-analysis. *Clin Infect Dis* 2015; 61(3): 418–426.
34. Latimer RL, Vodstrcil LA, Plummer EL, et al. The clinical indications for testing women for *Mycoplasma genitalium*. *Sex Transm Infect* 2022; 98(4): 277–285.
35. Bainbridge ED, Soge OO, Grabow C, et al. 163. High prevalence of urogenital and rectal *Mycoplasma genitalium* in U.S. MSM with a history of STIs in the last year. *Open Forum Infect Dis* 2021; 8: S98–S99.
36. Barbee L, Golden M, Soge O, et al. Rectal and pharyngeal *M. Genitalium* among men who have sex with men (MSM): results from a longitudinal cohort study. *Sex Transm Infect* 2019; 95: A258.
37. Latimer RL, Shilling HS, Vodstrcil LA, et al. Prevalence of *Mycoplasma genitalium* by anatomical site in men who have sex with men: a systematic review and meta-analysis. *Sex Transm Infect* 2020; 96(8): 563–570.
38. Foschi C, Gaspari V, Sgubbi P, et al. Sexually transmitted rectal infections in a cohort of ‘men having sex with men’. *J Med Microbiol* 2018; 67(8): 1050–1057.
39. Ong JJ, Ruan L, Lim AG, et al. Impact of screening on the prevalence and incidence of *Mycoplasma genitalium* and its macrolide resistance in men who have sex with men living in Australia: a mathematical model. *EClinicalMedicine* 2021; 33: 100779.
40. Chow EPF, Lee D, Bond S, et al. Nonclassical pathogens as causative agents of proctitis in men who have sex with men. *Open Forum Infect Dis* 2021; 8(7): ofab137.
41. Bjornelius E, Jensen JS and Lidbrink P. Conjunctivitis associated with *Mycoplasma genitalium* infection. *Clin Infect Dis* 2004; 39(7): e67–e69.
42. Simms I, Eastick K, Mallinson H, et al. Associations between *Mycoplasma genitalium*, *Chlamydia trachomatis*, and pelvic inflammatory disease. *Sex Transm Infect* 2003; 79(2): 154–156.
43. Dean G, Whetham J, Soni S, et al. O031 pelvic inflammatory disease (PID), *Mycoplasma genitalium* and macrolide resistance in England. *Sex Transm Infect* 2016; 92(Suppl 1): A12.
44. Lewis J, Horner PJ and White PJ. Incidence of pelvic inflammatory disease associated with *Mycoplasma genitalium* infection: evidence synthesis of cohort study data. *Clin Infect Dis* 2020; 71(10): 2719–2722.
45. Cohen CR, Manhart LE, Bukusi EA, et al. Association between *Mycoplasma genitalium* and acute endometritis. *Lancet* 2002; 359(9308): 765–766.
46. Taylor BD, Zheng X, O’Connell CM, et al. Risk factors for *Mycoplasma genitalium* endometritis and incident infection: a secondary data analysis of the T cell response against *Chlamydia* (TRAC) study. *Sex Transm Infect* 2018; 94(6): 414–420.
47. Baczynska A, Funch P, Fedder J, et al. Morphology of human Fallopian tubes after infection with *Mycoplasma genitalium* and *Mycoplasma hominis*-in vitro organ culture study. *Hum Reprod* 2007; 22(4): 968–979.
48. Wiesenfeld HC and Manhart LE. *Mycoplasma genitalium* in women: current knowledge and research priorities for this recently emerged pathogen. *J Infect Dis* 2017; 216(suppl\_2): S389–S395.
49. Ito S, Tsuchiya T, Yasuda M, et al. Prevalence of genital mycoplasmas and ureaplasmas in men younger than 40 years-of-age with acute epididymitis. *Int J Urol* 2012; 19(3): 234–238.
50. Fernandez Vecilla D, Aragon Diez J, Nieto Toboso MC, et al. Could *Mycoplasma genitalium* be involved in chronic

- granulomatous orchiepididymitis? Case report and literature review. *Rev Esp Quimioter* 2023; 36(1): 103–106.
51. Papeš D, Pasini M, Jerončić A, et al. Detection of sexually transmitted pathogens in patients with chronic prostatitis/chronic pelvic pain: a prospective clinical study. *Int J STD AIDS* 2017; 28(6): 613–615.
  52. Rossotti R, Travi G, Bana NB, et al. A case of chronic bacterial prostatitis due to *Mycoplasma genitalium*. *Sex Transm Infect* 2023; 99(8): 571–573.
  53. Taylor-Robinson D, Gilroy CB, Horowitz S, et al. *Mycoplasma genitalium* in the joints of two patients with arthritis. *Eur J Clin Microbiol Infect Dis* 1994; 13(12): 1066–1069.
  54. Simos P and Stewart AG. Sexually acquired reactive arthritis secondary to macrolide-resistant *Mycoplasma genitalium* urethritis. *Intern Med J* 2022; 52(2): 332–333.
  55. Frenzer C, Egli-Gany D, Vallely LM, et al. Adverse pregnancy and perinatal outcomes associated with *Mycoplasma genitalium*: systematic review and meta-analysis. *Sex Transm Infect* 2022; 98(3): 222–227.
  56. British Association for Sexual Health and HIV (BASHH). BASHH summary guidance on testing for sexually transmitted infections, 2023. [https://www.bashh.org/\\_userfiles/pages/files/resources/bashh\\_summary\\_guidance\\_on\\_stis\\_testing\\_2023.pdf](https://www.bashh.org/_userfiles/pages/files/resources/bashh_summary_guidance_on_stis_testing_2023.pdf)
  57. Unemo M, Bradshaw CS, Hocking JS, et al. Sexually transmitted infections: challenges ahead. *Lancet Infect Dis* 2017; 17(8): e235–e279.
  58. Le Roy C, Le Hen I, Clerc M, et al. The first performance report for the Bio-Rad Dx CT/NG/MG assay for simultaneous detection of *Chlamydia trachomatis*, *Neisseria gonorrhoeae* and *Mycoplasma genitalium* in urogenital samples. *J Microbiol Methods* 2012; 89(3): 193–197.
  59. Tabrizi SN, Tan LY, Walker S, et al. Multiplex assay for simultaneous detection of *Mycoplasma genitalium* and macrolide resistance using PlexZyme and PlexPrime technology. *PLoS One* 2016; 11(6): e0156740.
  60. Shipitsyna E, Zolotoverkhaya E, Dohn B, et al. First evaluation of polymerase chain reaction assays used for diagnosis of *Mycoplasma genitalium* in Russia. *J Eur Acad Dermatol Venereol* 2009; 23(10): 1164–1172.
  61. Jensen JS, Bjornelius E, Dohn B, et al. Comparison of first void urine and urogenital swab specimens for detection of *Mycoplasma genitalium* and *Chlamydia trachomatis* by polymerase chain reaction in patients attending a sexually transmitted disease clinic. *Sex Transm Dis* 2004; 31(8): 499–507.
  62. Gaydos CA, Manhart LE, Taylor SN, et al. Molecular testing for *Mycoplasma genitalium* in the United States: results from the AMES prospective multicenter clinical study. *J Clin Microbiol*. 2019; 57(11): 57.
  63. Dize L, Barnes P Jr, Barnes M, et al. Performance of self-collected penile-meatal swabs compared to clinician-collected urethral swabs for the detection of *Chlamydia trachomatis*, *Neisseria gonorrhoeae*, *Trichomonas vaginalis*, and *Mycoplasma genitalium* by nucleic acid amplification assays. *Diagn Microbiol Infect Dis* 2016; 86(2): 131–135.
  64. Jaya ZN, Mapanga W, Dlangalala T, et al. Accuracy of self-collected versus healthcare worker collected specimens for diagnosing sexually transmitted infections in females: an updated systematic review and meta-analysis. *Sci Rep* 2024; 14(1): 10496.
  65. Mobley VL, Hobbs MM, Lau K, et al. *Mycoplasma genitalium* infection in women attending a sexually transmitted infection clinic: diagnostic specimen type, coinfections, and predictors. *Sex Transm Dis* 2012; 39(9): 706–709.
  66. Lillis RA, Nsuami MJ, Myers L, et al. Utility of urine, vaginal, cervical, and rectal specimens for detection of *Mycoplasma genitalium* in women. *J Clin Microbiol* 2011; 49(5): 1990–1992.
  67. Van Der Pol B, Waites KB, Xiao L, et al. *Mycoplasma genitalium* detection in urogenital specimens from symptomatic and asymptomatic men and women by use of the cobas TV/MG Test. *J Clin Microbiol* 2020 May 26;58(6): e02124-19.
  68. Clinical Effectiveness Group of the British Association for Sexual Health and HIV (BASHH). Patient information leaflet - *Mycoplasma genitalium* 2021. [https://www.bashh.org/resources/59/mycoplasma\\_genitalium](https://www.bashh.org/resources/59/mycoplasma_genitalium)
  69. Lau A, Bradshaw CS, Lewis D, et al. The efficacy of azithromycin for the treatment of genital *Mycoplasma genitalium*: a systematic review and meta-analysis. *Clin Infect Dis* 2015; 61(9): 1389–1399.
  70. Chua T-P, Vodstrcil LA, Murray GL, et al. Evolving patterns of macrolide and fluoroquinolone resistance in *Mycoplasma genitalium*: an updated global systematic review and meta-analysis. SSRN. 2024. DOI: [10.2139/ssrn.4764790](https://doi.org/10.2139/ssrn.4764790).
  71. UK Health Security Agency. *Mycoplasma genitalium* antimicrobial resistance surveillance (MARS) report: 2023. <https://www.gov.uk/government/publications/mycoplasma-genitalium-antimicrobial-resistance-surveillance-mars/mycoplasma-genitalium-antimicrobial-resistance-surveillance-mars-report-2023>
  72. Durukan D, Read TRH, Murray G, et al. Resistance-guided antimicrobial therapy using doxycycline-moxifloxacin and doxycycline-2.5 g azithromycin for the treatment of *Mycoplasma genitalium* infection: efficacy and tolerability. *Clin Infect Dis* 2020; 71(6): 1461–1468.
  73. Read TRH, Fairley CK, Murray GL, et al. Outcomes of resistance-guided sequential treatment of *Mycoplasma genitalium* Infections: a prospective evaluation. *Clin Infect Dis* 2019; 68(4): 554–560.
  74. Gesink DC, Mulvad G, Montgomery-Andersen R, et al. *Mycoplasma genitalium* presence, resistance and epidemiology in Greenland. *Int J Circumpolar Health* 2012; 71: 1–8.
  75. Manhart LE, Jensen JS, Bradshaw CS, et al. Efficacy of antimicrobial therapy for *Mycoplasma genitalium* infections. *Clin Infect Dis* 2015; 61(Suppl 8): S802–S817.
  76. Murray G, Plummer EL, Bodihabadu K, et al. GyrA mutations in *Mycoplasma genitalium* contribute to moxifloxacin failure - a new target for resistance-guided therapy. *Sex Transm Dis* 2024; 51(1): S18.
  77. Medicines and Healthcare products Regulatory Agency. Fluoroquinolone antibiotics: must now only be prescribed when other commonly recommended antibiotics are inappropriate 2024. <https://www.gov.uk/drug-safety-update/fluoroquinolone-antibiotics-must-now-only-be-prescribed-when-other-commonly-recommended-antibiotics-are-inappropriate>

prescribed-when-other-commonly-recommended-antibiotics-are-inappropriate

78. Li Y, Le WJ, Li S, et al. Meta-analysis of the efficacy of moxifloxacin in treating Mycoplasma genitalium infection. *Int J STD AIDS* 2017; 28(11): 1106–1114.
79. Clarke EJ, Vodstrcil LA, Plummer EL, et al. Efficacy of minocycline for the treatment of Mycoplasma genitalium. *Open Forum Infect Dis* 2023; 10(8): ofad427.
80. Hamasuna R, Jensen JS and Osada Y. Antimicrobial susceptibilities of Mycoplasma genitalium strains examined by broth dilution and quantitative PCR. *Antimicrob Agents Chemother* 2009; 53(11): 4938–4939.
81. Read TRH, Jensen JS, Fairley CK, et al. Use of pristinamycin for macrolide-resistant Mycoplasma genitalium infection. *Emerg Infect Dis* 2018; 24(2): 328–335.
82. Htaik K, Vodstrcil LA, Plummer EL, et al. Efficacy and tolerability of the combination of minocycline and metronidazole for macrolide-resistant Mycoplasma genitalium. *J Antimicrob Chemother* 2025; 80: 1878–1884.
83. Perin J, Coleman JS, Ronda J, et al. Maternal and fetal outcomes in an observational cohort of women with Mycoplasma genitalium infections. *Sex Transm Dis* 2021; 48(12): 991–996.
84. Bar-Oz B, Weber-Schoendorfer C, Berlin M, et al. The outcomes of pregnancy in women exposed to the new macrolides in the first trimester: a prospective, multicentre, observational study. *Drug Saf* 2012; 35(7): 589–598.
85. Sarkar M, Woodland C, Koren G, et al. Pregnancy outcome following gestational exposure to azithromycin. *BMC Pregnancy Childbirth* 2006; 6: 18.
86. Cooper WO, Hernandez-Diaz S, Arbogast PG, et al. Antibiotics potentially used in response to bioterrorism and the risk of major congenital malformations. *Paediatr Perinat Epidemiol* 2009; 23(1): 18–28.
87. British Association for Sexual Health and HIV (BASHH). BASHH position statement on doxycycline use in pregnancy, 2024. [https://www.bashh.org/resources/101/bashh\\_position\\_statement\\_on\\_doxycycline\\_use\\_in\\_pregnancy](https://www.bashh.org/resources/101/bashh_position_statement_on_doxycycline_use_in_pregnancy)
88. Moxifloxacin - summary of product characteristics (SmPC). updated 24 July 2008. Available from: [https://www.ema.europa.eu/en/documents/referral/avelox-article-6-12-referral-annex-i-ii-iii\\_en.pdf](https://www.ema.europa.eu/en/documents/referral/avelox-article-6-12-referral-annex-i-ii-iii_en.pdf)
89. Minocycline - summary of product characteristics (SmPC). updated 15 July 2024. Available from: <https://www.medicines.org.uk/emc/product/9519/smpc/print>
90. Kelsey JJ, Moser LR, Jennings JC, et al. Presence of azithromycin breast milk concentrations: a case report. *Am J Obstet Gynecol* 1994; 170(5 Pt 1): 1375–1376.
91. Lund M, Pasternak B, Davidsen RB, et al. Use of macrolides in mother and child and risk of infantile hypertrophic pyloric stenosis: nationwide cohort study. *BMJ* 2014; 348: g1908.
92. Doxycycline In: Joint Formulary Committee. *British national formulary*. London: British Medical Association and Royal Pharmaceutical Society of Great Britain. Available from: <https://bnf.nice.org.uk/drugs/doxycycline/>
93. Compendium EM. *Doxycycline 100mg capsules- summary of product characteristics (SmPC)*. UK: Datapharm, 2021. [cited 2025 April 3]. Available from: <https://www.medicines.org.uk/emc/product/13082/smpc> (accessed 3 April 2025).
94. *Pyostacine - summary of product characteristics (SmPC)*. Paris, France: Sanofi-Aventis Paris, France; 2012. updated 12 July 2012. Available from: [https://www.nhstaysideadtc.scot.nhs.uk/Antibiotic\\_site/pdf\\_docs/Pyostacine\\_SPC.pdf](https://www.nhstaysideadtc.scot.nhs.uk/Antibiotic_site/pdf_docs/Pyostacine_SPC.pdf)
95. Kong FYS, Horner P, Unemo M, et al. Pharmacokinetic considerations regarding the treatment of bacterial sexually transmitted infections with azithromycin: a review. *J Antimicrob Chemother* 2019; 74(5): 1157–1166.
96. Ito S, Shimada Y, Yamaguchi Y, et al. Selection of Mycoplasma genitalium strains harbouring macrolide resistance-associated 23S rRNA mutations by treatment with a single 1 g dose of azithromycin. *Sex Transm Infect* 2011; 87(5): 412–414.
97. Manhart LE, Gillespie CW, Lowens MS, et al. Standard treatment regimens for nongonococcal urethritis have similar but declining cure rates: a randomized controlled trial. *Clin Infect Dis* 2013; 56(7): 934–942.
98. Falk L, Enger M and Jensen JS. Time to eradication of Mycoplasma genitalium after antibiotic treatment in men and women. *J Antimicrob Chemother* 2015; 70(11): 3134–3140.
99. Slifirski JB, Vodstrcil LA, Fairley CK, et al. Mycoplasma genitalium Infection in adults reporting sexual contact with infected partners, Australia, 2008-2016. *Emerg Infect Dis* 2017; 23(11): 1826–1833.
100. Cina M, Baumann L, Egli-Gany D, et al. Mycoplasma genitalium incidence, persistence, concordance between partners and progression: systematic review and meta-analysis. *Sex Transm Infect* 2019; 95(5): 328–335.
101. McClean H, Radcliffe K, Sullivan A, et al. 2012 BASHH statement on partner notification for sexually transmissible infections. *Int J STD AIDS* 2013; 24(4): 253–261.

## Appendix

### Abbreviations

AGREE II	Appraisal of Guidelines, Research and Evaluation
AMR	Antimicrobial Resistance
ASHM	Australasian Society for HIV, Viral Hepatitis and Sexual Health Medicine
BASHH	British Association for Sexual Health and HIV
bd	Twice Daily
BHIVA	British HIV Association
BNF	British National Formulary
CEG	Clinical Effectiveness Group
CI	Confidence Interval
ECCMID	European Society of Clinical Microbiology and Infectious Diseases
EQI	Equality Impact Assessment
FDA	Food and Drug Administration
GBMSM	Gay, Bisexual and other Men who have sex with men
GRADE	Grading of Recommendations, Assessment, Development, and Evaluations

GUMCAD	Genitourinary Medicine Clinic Activity Dataset	NGU	Non-gonococcal Urethritis
HIV	Human Immunodeficiency Virus	NHS	National Health Service
ICAAC	Interscience Conference of Antimicrobial Agents and Chemotherapy	od	Once Daily
IUSTI	International Union against Sexually Transmitted Infection, BASHH	OR	Odds Ratio
MHRA	Medicines and Healthcare Products Regulatory Agency	PID	Pelvic Inflammatory Disease
MIC	Minimum Inhibitory Concentration	PrEP	Pre-exposure Prophylaxis
MRAM	Macrolide Resistance Associated Mutations	SHHAPT	Sexual Health and HIV Activity Property Type
MSW	Men who have sex with Women	SmPC	Summary of Product Characteristics
NAAT	Nucleic Acid Amplification Test	STI	Sexually Transmitted Infection
NCNGU	Non-chlamydial Non-gonococcal Urethritis	TOC	Test of Cure
		UK	United Kingdom
		UKHSA	UK Health Security Agency
		UKTIS	UK Teratology Information Services.